

The first total synthesis of lymphostin

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Abstract—Lymphostin (**1**), a novel immunosuppressant, has been synthesized from tryptophan through six kinds of regioselectively oxidative reactions.

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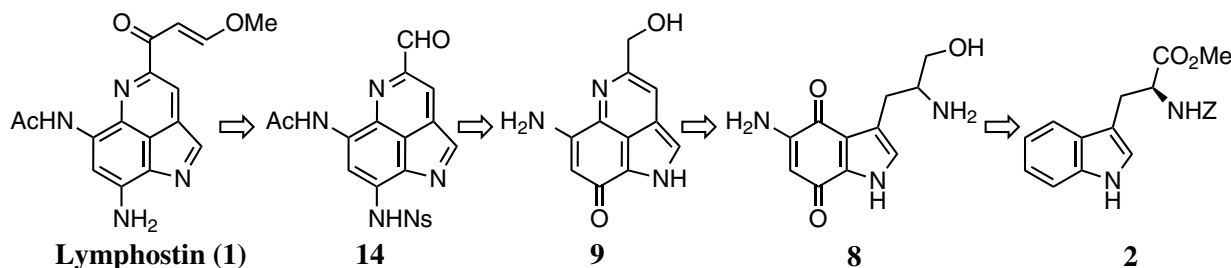
Lymphostin (**1**) was isolated by the Kyowa Hakko group from a culture broth of *Streptomyces* sp. as an immunosuppressant to show potent inhibitory activity against lymphocyte kinase.¹ The structure was elucidated by thorough NMR studies to be a novel tricyclic aromatic alkaloid with a pyrroloquinoline skeleton.² In addition to its novel mechanism of action, impinging on a crucial biological cascade, the structure of **1** interested us as a focused target for total synthesis. We have proposed lymphostin (**1**) to be biogenetically produced from tryptophan.

Herein, we report the implementation of a novel biomimetic strategy for the first total synthesis of lymphostin (**1**). The critical element of the synthetic plan was inspired by our proposed biosynthetic sequence as shown in the retrosynthetic perspective to assemble the tricyclic core **14** from the quinone **8** through cyclization and oxidation to give the intermediary iminoquinone **9** (Scheme 1).

In the event, the starting *N*-carbobenzyloxy-L-tryptophan methyl ester (**2**) was converted to the racemic ketoalcohol **3** in a quantitative yield by hydride reduction and DDQ oxidation (Scheme 2 and Table 1). The selective introduction of a hydroxyl group onto the phenyl portion proceeded on oxidative reaction with $\text{Ti}(\text{OCOCF}_3)_3$ followed by treatment with $\text{CuSO}_4 \cdot 5\text{H}_2\text{O}$ to give the phenol **4**.^{3,4} This was deoxygenated by NaBH_3CN to give **5**,⁵ which was oxidized by $\text{NO}(\text{SO}_3\text{K})_2$ to the quinone **6** in high yield.

Regioselective reaction of **6** with benzylamine to afford **7**⁶ was followed by hydrogenolysis to the diamino derivative **8**. This was cyclized concomitant with aromatization by heating in DMF with air to give the desired iminoquinone **9**, which was acetylated to **10**.⁷

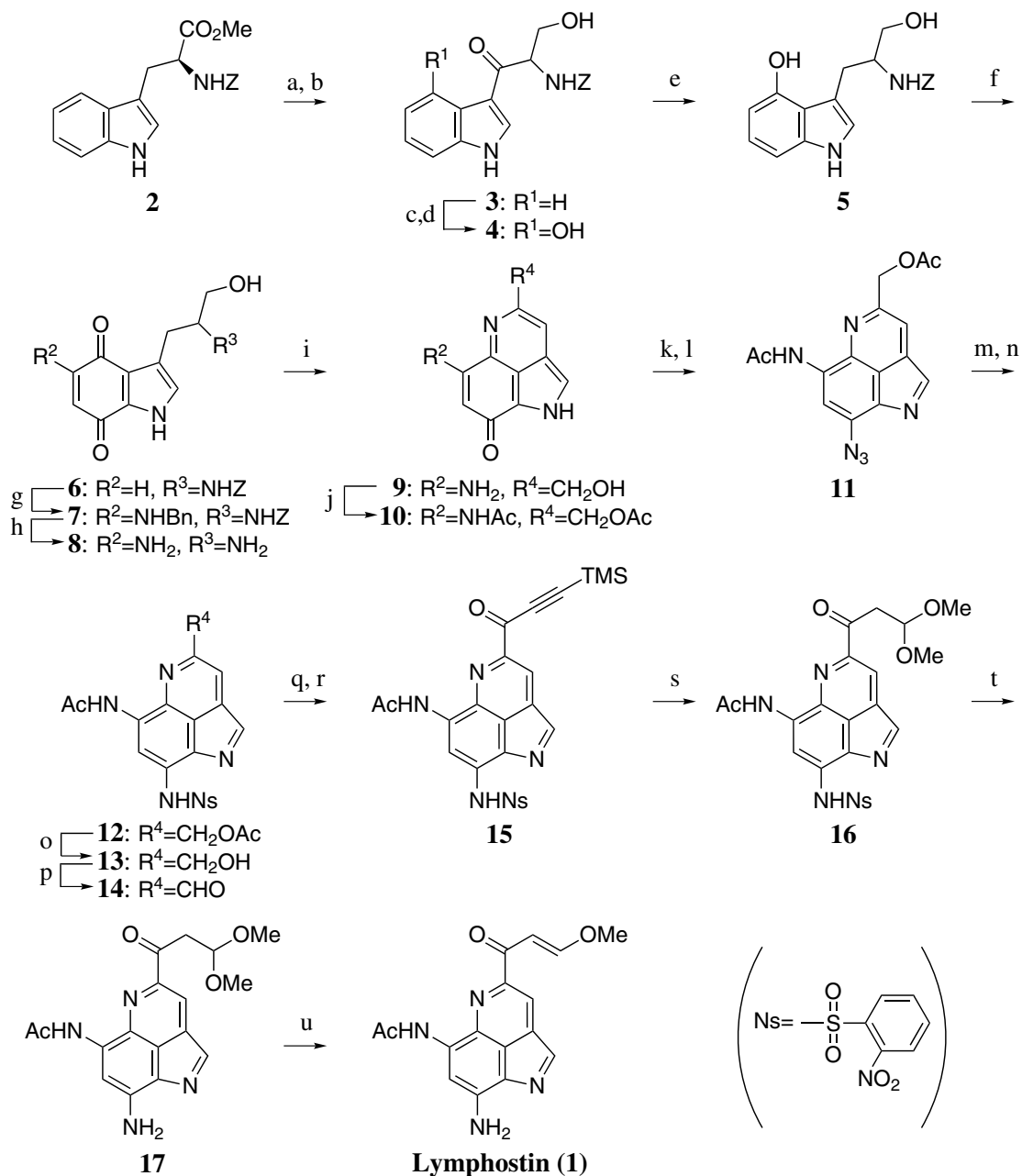
The transformation of the quinone **10** to the diamino-pyrroloquinoline **12** was examined under a variety of conditions, and realized by the introduction of an azido



Scheme 1.

Keywords: Total synthesis; Immunosuppressant; Pyrroloquinoline; Tricyclic alkaloid.

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Scheme 2. Reagents and conditions: (a) LiBH₄/THF, rt, 1 h then reflux, 1 h; (b) DDQ/aq THF, rt, 1 h, 95% in two steps; (c) Ti(OCOCF₃)₃/TFA, rt, 1 h; (d) CuSO₄·5H₂O/DMF, 130 °C, 10 min, 59% in two steps; (e) NaBH₃CN, BF₃·Et₂O/THF, rt, 4 h, 85%; (f) NO(SO₃K)₂/acetone-pH 7 phosphate buffer, rt, 2 h, 82%; (g) BnNH₂, air/PhH, rt, 4 h, 79%; (h) H₂, Pd-C/EtOH, rt, 2 h, quant.; (i) air/DMF, 70 °C, 1 h; (j) C₆F₅OAc/DMF, rt, 12 h, 66% in two steps; (k) Tf₂O, Py/CH₂Cl₂, -78 °C, 1 h; (l) *n*-Bu₄NN₃/CH₂Cl₂, -78 °C to rt, 12 h, 68% in two steps; (m) H₂, Lindlar cat./EtOH, rt, 2 h; (n) NsCl, Py/ClCH₂CH₂Cl, 50 °C, 1 h, 75% in two steps; (o) aq acetone, rt, 5 days, quant.; (p) PCC, Al₂O₃/AcOH/CH₂Cl₂, rt, 4 h, 80%; (q) trimethylsilylacetylene, *n*-BuLi/THF, -78 °C, 2 h; (r) MnO₂/acetone/CH₂Cl₂, rt, 12 h, 69% in two steps; (s) K₂CO₃/MeOH, rt, 5 h, 73%; (t) PhSH, K₂CO₃/DMF, rt, 12 h, 75%; (u) MS 4A/DMSO, 90 °C, 7 days, 64%.

group, and subsequent reduction. Treatment of **10** with Tf₂O followed by *n*-Bu₄NN₃ afforded the tautomeric azide **11**, which was reduced on the Lindlar catalyst and protected by an *o*-nitrobenzenesulfonyl (Ns) group⁸ to afford **12**. De-O-acetylation of **12** to **13** was quantitatively carried out only with aqueous acetone. The alcohol **13** was oxidized to the aldehyde **14**, which was followed successively by reaction with trimethylsilylacetylene and oxidation with MnO₂ to give the ethynyl ketone **15**.⁹ This was transformed to the dimethyl acetal

16 by alkaline methanol and de-N-protected by alkaline thiophenol to the amine **17**.

In the final stage, methanol was eliminated from **17**¹⁰ to give lymphostin (**1**). This was identical in all respects with the natural product,¹¹ completing the total synthesis.

At the heart of the synthesis of lymphostin (**1**) described herein are six kinds of oxidative reactions, which were regioselectively accomplished.

Table 1. Physico-chemical properties of compounds

Compounds	Mp (°C)	¹ H NMR (600 MHz; δ ppm; <i>J</i> Hz) ¹³ C NMR (150 MHz; δ ppm) MS (<i>m/z</i>) IR (KBr; cm ⁻¹)	Compounds	Mp (°C)	¹ H NMR (600 MHz; δ ppm; <i>J</i> Hz) ¹³ C NMR (150 MHz; δ ppm) MS (<i>m/z</i>) IR (KBr; cm ⁻¹)
1	276–277 Crystals (<i>i</i> -PrOH)	¹ H NMR (DMSO- <i>d</i> ₆): δ 2.35 (3H, s), 3.95 (3H, s), 7.64 (1H, d, <i>J</i> = 12.5 Hz), 7.88 (1H, d, <i>J</i> = 12.5 Hz), 8.15 (1H, s), 8.23 (2H, br s), 8.38 (1H, s), 8.63 (1H, s), 10.16 (1H, br s) ¹³ C NMR (DMSO- <i>d</i> ₆): δ 24.3, 58.4, 101.7, 110.6, 117.3, 124.3, 124.8, 133.5, 136.4, 139.5, 145.6, 147.5, 150.1, 163.8, 169.5, 188.5 FAB-MS: 311 [M+H] ⁺ IR (KBr): 3448, 1685, 1654, 1632, 1596, 1500, 1363, 1350, 1241, 1132, 1078	11	Wax	¹ H NMR (MeOH- <i>d</i> ₄): δ 2.15 (3H, s), 2.33 (3H, s), 5.47 (2H, s), 7.78 (1H, s), 8.03 (1H, s), 8.06 (1H, s) MALDI-TOF-MS: 325 [M+H] ⁺ IR (KBr): 3428, 2346, 1735, 1658, 1560, 1540, 1457, 1421, 1388, 1236, 1218, 1106, 1087
3	153 Needles (EtOAc)	¹ H NMR (MeOH- <i>d</i> ₄): δ 3.84 (1H, dd, <i>J</i> = 11.5 & 6.0 Hz), 3.96 (1H, dd, <i>J</i> = 11.5 & 5.0 Hz), 5.09 (1H, d, <i>J</i> = 13.0 Hz), 5.11 (1H, dd, <i>J</i> = 6.0 & 5.0 Hz), 5.12 (1H, d, <i>J</i> = 13.0 Hz), 7.21 (1H, ddd, <i>J</i> = 7.5, 7.5 & 1.0 Hz), 7.24 (1H, ddd, <i>J</i> = 7.5, 7.5 & 1.0 Hz), 7.27–7.38 (5H, m), 7.45 (1H, ddd, <i>J</i> = 7.5, 1.0 & 0.5 Hz), 8.25 (1H, ddd, <i>J</i> = 7.5, 1.0 & 0.5 Hz) FAB-MS: 339 [M+H] ⁺	12	218 (decomp.) Needles (MeOH)	¹ H NMR (MeOH- <i>d</i> ₄): δ 2.09 (3H, s), 2.28 (3H, s), 5.36 (2H, s), 7.80 (1H, ddd, <i>J</i> = 8.0, 8.0 & 1.0 Hz), 8.02 (1H, dd, <i>J</i> = 8.0 & 1.0 Hz), 8.05 (1H, dd, <i>J</i> = 8.0 & 1.0 Hz), 8.10 (1H, s), 8.11 (1H, ddd, <i>J</i> = 8.0, 8.0 & 1.0 Hz), 8.14 (1H, s), 8.26 (1H, s) FAB-MS: 484 [M+H] ⁺
4	196–197 Crystals (EtOAc)	¹ H NMR (MeOH- <i>d</i> ₄): δ 3.83 (1H, dd, <i>J</i> = 11.0 & 6.5 Hz), 3.95 (1H, dd, <i>J</i> = 11.0 & 6.5 Hz), 5.09 (1H, d, <i>J</i> = 14.0 Hz), 5.13 (1H, d, <i>J</i> = 14.0 Hz), 5.17 (1H, dd, <i>J</i> = 6.5 & 6.5 Hz), 6.55 (1H, d, <i>J</i> = 8.0 Hz), 6.91 (1H, d, <i>J</i> = 8.0 Hz), 7.12 (1H, dd, <i>J</i> = 8.0 & 8.0 Hz), 7.25–7.37 (5H, m), 8.31 (1H, s) FAB-MS: 355 [M+H] ⁺	13	225 (decomp.) Needles (EtOAc)	¹ H NMR (DMSO- <i>d</i> ₆): δ 2.16 (1H, br s), 2.20 (3H, s), 5.50 (2H, s), 7.88 (1H, ddd, <i>J</i> = 8.0, 8.0 & 1.0 Hz), 8.00 (1H, br s), 8.07 (1H, dd, <i>J</i> = 8.0 & 1.0 Hz), 8.08 (1H, s), 8.10 (1H, dd, <i>J</i> = 8.0 & 1.0 Hz), 8.11 (1H, s), 8.22 (1H, s), 8.23 (1H, ddd, <i>J</i> = 8.0, 8.0 & 1.0 Hz), 10.03 (1H, br s) MALDI-TOF-MS: 442 [M+H] ⁺
6	165 (decomp.) Prisms (acetone)	¹ H NMR (MeOH- <i>d</i> ₄): δ 2.81 (1H, dd, <i>J</i> = 14.5 & 10.0 Hz), 3.04 (1H, dd, <i>J</i> = 14.5 & 5.0 Hz), 3.56 (1H, dd, <i>J</i> = 11.0 & 5.0 Hz), 3.59 (1H, dd, <i>J</i> = 11.0 & 5.5 Hz), 3.92 (1H, dddd, <i>J</i> = 10.0, 5.5, 5.0 & 5.0 Hz), 4.92 (1H, d, <i>J</i> = 11.5 Hz), 5.03 (1H, d, <i>J</i> = 11.5 Hz), 6.50 (1H, d, <i>J</i> = 10.5 Hz), 6.53 (1H, d, <i>J</i> = 10.5 Hz), 6.96 (1H, s), 7.19–7.31 (5H, m) FAB-MS: 355 [M+H] ⁺	14	194–195 (decomp.) Amorphous	¹ H NMR (MeOH- <i>d</i> ₄): δ 2.18 (3H, s), 7.85 (1H, ddd, <i>J</i> = 8.0, 8.0 & 1.5 Hz), 7.90 (1H, dd, <i>J</i> = 7.5 & 1.5 Hz), 8.07 (1H, s), 8.09 (1H, s), 8.13 (1H, dd, <i>J</i> = 8.0 & 1.0 Hz), 8.16 (1H, ddd, <i>J</i> = 8.0, 7.5 & 1.0 Hz), 8.25 (1H, s), 10.03 (1H, s) FAB-MS: 440 [M+H] ⁺
7	>300 Crystals (MeOH)	¹ H NMR (MeOH- <i>d</i> ₄): δ 2.79 (1H, dd, <i>J</i> = 12.0 & 10.0 Hz), 3.01 (1H, dd, <i>J</i> = 12.0 & 6.0 Hz), 3.56 (1H, dd, <i>J</i> = 12.5 & 6.0 Hz), 3.57 (1H, dddd, <i>J</i> = 10.0, 6.0, 6.0 & 2.5 Hz), 3.60 (1H, dd, <i>J</i> = 12.5 & 2.5 Hz), 4.39 (2H, s), 4.93 (1H, d, <i>J</i> = 13.0 Hz), 5.04 (1H, d, <i>J</i> = 13.0 Hz), 5.08 (1H, s), 6.75 (1H, s), 7.16–7.32 (10H, m) FAB-MS: 460 [M+H] ⁺	15	244–246 (decomp.) Amorphous	¹ H NMR (MeOH- <i>d</i> ₄): δ 0.06 (9H, s), 2.23 (3H, s), 8.03 (1H, ddd, <i>J</i> = 8.0, 8.0 & 1.0 Hz), 8.06 (1H, s), 8.08 (1H, dd, <i>J</i> = 8.0 & 1.0 Hz), 8.12 (1H, dd, <i>J</i> = 8.0 & 1.0 Hz), 8.15 (1H, s), 8.17 (1H, ddd, <i>J</i> = 8.0, 8.0 & 1.0 Hz), 8.30 (1H, s) FAB-MS: 536 [M+H] ⁺
8	216–218 (decomp.) Crystals (MeOH)	¹ H NMR (MeOH- <i>d</i> ₄): δ 2.87 (1H, dd, <i>J</i> = 14.5 & 7.5 Hz), 2.96 (1H, dd, <i>J</i> = 14.5 & 7.0 Hz), 3.42 (1H, dddd, <i>J</i> = 7.5, 7.0, 6.5 & 4.0 Hz), 3.49 (1H, dd, <i>J</i> = 11.5 & 6.5 Hz), 3.68 (1H, dd, <i>J</i> = 11.5 & 4.0 Hz), 5.39 (1H, s), 6.83 (1H, s) ¹³ C NMR (MeOH- <i>d</i> ₄): δ 28.2, 54.4, 63.6, 98.2, 120.2, 121.3, 123.9, 136.5, 154.1, 180.1, 180.4 FAB-MS: 236 [M+H] ⁺	16	>300 Amorphous	¹ H NMR (MeOH- <i>d</i> ₄): δ 2.21 (3H, s), 3.28 (6H, s), 3.72 (2H, d, <i>J</i> = 6.0 Hz), 4.93 (1H, t, <i>J</i> = 6.0 Hz), 7.98 (1H, ddd, <i>J</i> = 8.0, 7.5 & 1.0 Hz), 8.06 (1H, s), 8.12 (1H, dd, <i>J</i> = 8.0 & 1.0 Hz), 8.14 (1H, s), 8.15 (1H, dd, <i>J</i> = 8.0 & 1.0 Hz), 8.20 (1H, ddd, <i>J</i> = 8.0, 7.5 & 1.0 Hz), 8.29 (1H, s) FAB-MS: 528 [M+H] ⁺

Table 1 (continued)

Com-pounds	Mp (°C)	¹ H NMR (600 MHz; δ ppm; J Hz) ¹³ C NMR (150 MHz; δ ppm) MS (m/z) IR (KBr; cm ⁻¹)	Com-pounds	Mp (°C)	¹ H NMR (600 MHz; δ ppm; J Hz) ¹³ C NMR (150 MHz; δ ppm) MS (m/z) IR (KBr; cm ⁻¹)
10	182 (decomp.) Amorphous	¹ H NMR (MeOH- <i>d</i> ₄): δ 2.16 (3H, s), 2.33 (3H, s), 5.43 (2H, s), 7.71 (1H, s), 7.99 (1H, s), 8.02 (1H, s) ¹³ C NMR (DMSO- <i>d</i> ₆): δ 21.1, 24.9, 67.3, 117.6, 118.1, 119.9, 120.0, 124.0, 125.0, 140.8, 142.3, 147.3, 170.4, 170.6, 173.6 MALDI-TOF-MS: 300 [M+H] ⁺ IR (KBr): 3442, 1739, 1685, 1592, 1508	17	288–290 (decomp.) Amorphous	¹ H NMR (DMSO- <i>d</i> ₆): δ 2.33 (3H, s), 3.32 (6H, s), 3.78 (2H, d, J = 5.5 Hz), 5.05 (1H, t, J = 5.5 Hz), 8.21 (1H, s), 8.30 (2H, br s), 8.52 (1H, s), 10.05 (1H, br s) FAB-MS: 343 [M+H] ⁺

It seems likely that the chemistry and insights developed for this total synthesis will find wider application.

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References and notes

- Nagata, H.; Ochiai, K.; Aotani, Y.; Ando, K.; Yoshida, M.; Takahashi, I.; Tamaoki, T. *J. Antibiot.* **1997**, *50*, 537.
- Aotani, Y.; Nagata, H.; Yoshida, M. *J. Antibiot.* **1997**, *50*, 543.
- Kogan, T. P.; Somers, T. C.; Venuti, M. C. *Tetrahedron* **1990**, *46*, 6623.
- Somei, M. *Yakugaku Zassi* **1988**, *108*, 361.
- Srikrishna, A.; Sattigeri, J. A.; Viswajanani, R.; Yelamagad, C. V. *Synlett* **1995**, 93.
- Jackson, Y. A.; Billimoria, A. D.; Sadanandan, E. V.; Cava, M. P. *J. Org. Chem.* **1995**, *60*, 3543.
- Kisfaludy, L.; Mohacsi, T.; Low, M.; Drexler, F. *J. Org. Chem.* **1979**, *44*, 654.
- Fukuyama, T.; Jow, C.-K.; Cheung, M. *Tetrahedron Lett.* **1995**, *36*, 6373.
- Takao, K.; Nagata, S.; Kobayashi, S.; Ito, H.; Taguchi, T. *Chem. Lett.* **1998**, 447.
- Akama, T.; Nagata, H.; Hasegawa, A.; Ue, H.; Takahashi, I.; Saitoh, Y.; Mochida, K.; Ikeda, S.; Kanda, Y. WO 00/02879, 2000.
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